SIGNIFICANCE OF LECITHIN:CHOLESTEROL ACYLTRANSFERASE ACTIVITY AS A PROGNOSTIC INDICATOR OF EARLY ALLOGRAFT FUNCTION IN CLINICAL LIVER TRANSPLANTATION¹,²

Mitsuo Shimada,² ³ Katsuhiko Yanaga,² Leonard Makowka,³ Saburo Kakizoe,⁴ David H. Van Thiel,⁵ and Thomas E. Starzl² ³

The Departments of Surgery, Pathology, and Medicine, University of Pittsburgh, School of Medicine, Pittsburgh, Pennsylvania 15261

University of Wisconsin (UW) solution by Belzer et al. (7, 8) for preservation of the liver has improved the results even further, and has allowed the procedure to be performed more widely. However, graft nonfunction—either primary, or secondary due to technical failure—remains a major complication of OLTx, and one that carries a high morbidity, mortality, and need for retransplantation (9). In an earlier study (10), we have demonstrated that the most commonly used donor characteristics did not distinguish between grafts that functioned and those that failed to function. Moreover, only a few studies have been reported on allograft viability in the early postoperative period (11–16).

Lecithin:cholesterol acyltransferase (LCAT), an enzyme that esterifies free cholesterol (17) and functions in the transportation of cholesterol and the metabolism of lipoproteins (18), has been known to be useful in assessing liver function following hepatic resection or other surgical procedures such as operations for esophageal varices (19). LCAT activity also has been shown to be a valuable indicator of prognosis in fulminant liver failure (20). Reduced LCAT activity in patients with liver diseases is considered to be due to impaired protein synthesis occurring secondary to hepatic dysfunction. Based upon the data obtained in various types of hepatic diseases, the present study was performed to determine if serial quantitation of the plasma LCAT activity could be utilized as a predictor of allograft function early after OLTx.

MATERIALS AND METHODS

During a three-month period between September 1 and November 30, 1987, 76 OLTx were performed in 68 adult patients at the Presbyterian University Hospital, University of Pittsburgh. The first 42 grafts were preserved in Euro-Collins' solution (4°C), while the other 34 were stored in University of Wisconsin (UW) solution (4°C). Eleven of these grafts (15%) exhibited poor function, as manifested by death due to primary nonfunction (two grafts), need for retransplantation within the first week after OLTx (4 grafts), and poor synthetic function with a peak prothrombin time of over 20 sec (control 11.8 sec) during the first 5 posttransplant days (5 grafts).

From the recipients of 9 of the 11 grafts with poor function, blood samples were obtained at predetermined time points: immediately prior to OLTx, immediately after OLTx; at 6, 12, 18, and 24 hr after OLTx; and at 3 days posttransplant. Plasma was separated from the blood by centrifugation at 2000 RPM for 5 min at 37°C and stored at –60°C. The selection was random and was based primarily on the availability of pretransplant blood samples. Plasma LCAT activity and pretransplant clinical variables of these recipients were compared with those of 15 control patients whose graft exhibited good function early after OLTx.

Since the first human orthotopic liver transplant (OLTx) performed in 1963, enormous progress has been made in the overall management and survival of patients with end-stage liver diseases (1–3). Factors responsible for this improved outcome include the introduction of cyclosporine (4), the use of venous-venous bypass (5), and a standardization of the technique of biliary tract reconstruction (6). The recent introduction of

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³ Department of Surgery.
⁴ Address correspondence to: Mitsuo Shimada, M.D., Department of Surgery II, Faculty of Medicine, Kyushu University, 3-1-1 Maidashi, Higashi-ku, Fukuoka, 812, Japan.
⁵ Department of Pathology.
⁶ Department of Medicine.
⁷ Address reprint requests to Thomas E. Starzl, M.D., Department of Surgery, University of Pittsburgh, Falk Clinic 4 West, 3601 Fifth Avenue, Pittsburgh, PA 15213.
tion (control), as manifested by a peak SGOT, SGPT, and prothrombin time of less than 2000 IU/L, 2000 IU/L, and 20 sec, respectively.

Plasma lecithin:cholesterol acyltransferase activity was determined using a LCAT test kit-S obtained from Nippon Shoji Co., Ltd., Osaka, Japan using a modification of the method of Nagasaki and Akanuma (21). The modification consists of the addition of ascorbic acid oxidase and sodium azide (NaN₃) to the mixture to minimize interference by plasma levels of ascorbic acid and hemoglobin, and also the use of DAO₆ (Sodium N-ethyl-N [2-hydroxy-3-sulfopropyl]-3,5-dimethoxyaniline) instead of phenol for the color reaction such that absorbance is read at 600 nm, thereby preventing the influence of the presence of bilirubin. Plasma LCAT activity was determined by its ability to esterify cholesterol at 37°C. A unit of LCAT activity was expressed as the reduction in free cholesterol (µg/ml/hr) in the assay mixture.

In order to assess the association between plasma LCAT activity and the quality of the allograft, histological findings of the grafts with poor function and the controls together were correlated with the LCAT activities. Allograft biopsy was obtained by a Trucut biopsy needle (Travenol Laboratories) immediately prior to implantation in 6 of the 11 grafts with poor function and all 15 control livers. The biopsy specimens were fixed with 10% buffered formalin and stained with hematoxylin and eosin. The histological findings were graded as follows:

Grade I (12 grafts)—minimal or no evidence of preservation damage;
Grade II (9 grafts)—changes consistent with ischemic injury, as manifested by diffuse hepatic cellular swelling, steatosis, or acidophilic necrosis.

For statistical analysis, the Mann-Whitney U and chi-square tests were used. Results of the plasma LCAT activity were expressed as means ± SD.

**RESULTS**

Table 1 demonstrates the relationship between LCAT activity during the first 24 hr after OLTx and graft function. A mean LCAT activity of less than 5 U correlated with poor graft function (P<0.05, χ² = 5.23). Table 2 lists the pre- and intraoperative clinical variables of the patients in each group. No significant difference was identified among the clinical parameters between the two groups. Similarly, preengraftment LCAT activity for both groups was not different. Table 3 lists the clinical data of the 9 patients who exhibited poor graft function. Three of these 9 patients (cases 6, 7, and 8) required retransplantation and one (case 1) died within a week.

Figure 1 and Table 4 depict the short-term changes of the LCAT activity after OLTx. Serial LCAT activity in the group with poor liver function was significantly lower than that of the group with adequate liver function at 6, 12, and 18 hr and at 3 days following OLTx (P<0.05 at most). Histologically, 12 grafts with pretransplant histology grade I had a mean LCAT activity within 24 hr after OLTx of 8.5±4.4 U, which was significantly higher than 4.6±1.9 U of 9 grafts with histology grade II (P<0.05).

**DISCUSSION**

This study demonstrated a statistically significant correlation between early allograft function and the plasma LCAT activity in OLTx recipients during the early postoperative period.

Numerous previous attempts have been made to assess the viability of the hepatic allograft prior to and immediately after transplantation. Fath et al. (11) reported a correlation between the ability of the hepatic allograft to reduce abnormal levels of total plasma amino acids immediately following revascularization and subsequent postoperative morbidity, defined as reoperation, death, or hepatic necrosis during the first 48 hr after OLTx. Jenkins et al. (12) demonstrated that a delayed recovery of the rate of central plasma clearance of amino acids following liver transplantation identifies recipients with an increased mortality. Taki et al. (13) reported in pigs that a failure to maintain a normal ratio of ketone bodies after OLTx correlated with hepatic energy charge and predicted a high mortality. Kamiike et al. (14) recently suggested that in humans the recovery of ATP levels in liver may be an indicator of early allograft function. Mora et al. (15) have demonstrated that the clearance of serum total bile acids can also serve as a putative marker of early allograft function (15). Persson et al. (16) have suggested that total-body oxygen consumption, which rapidly declines during the anhepatic phase of the transplant procedure and returns quickly to baseline levels immediately after revascularization of the graft, may also be a good indicator of early graft function and perfusion. All of these methods, however, are sophisticated, cumbersome, or invasive, or they require special equipment and expertise, and so cannot be used routinely and more widely.

Currently, measurement of the prothrombin time and determination of the levels of specific coagulation factors remain the most practical methods for the assessment of early allograft function after OLTx. Clinical features of malfunction of the liver allograft after OLTx include inability to awaken from anesthesia, low bile output or production of thin bile through a T tube, elevated transaminases and prothrombin time, and persistent or progressive hyperbilirubinemia, often followed by a rapid development of renal failure and other metabolic derangements. Nevertheless, as exhibited in Table 3, considerable

**Table 1. Correlation between allograft function and mean plasma LCAT activity during the first 24 hours following OLTx**

<table>
<thead>
<tr>
<th>Mean LCAT* activity (U)</th>
<th>Adequate</th>
<th>Poor</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;5*</td>
<td>3 (20.0%)</td>
<td>6 (66.7%)</td>
</tr>
<tr>
<td>≥5*</td>
<td>12 (80.0%)</td>
<td>3 (33.3%)</td>
</tr>
</tbody>
</table>

*LCAT, lecithin:cholesterol acyltransferase; OLTx, orthotopic liver transplantation.

**Table 2. Relationship between pre- or intraoperative clinical variables and early allograft function**

<table>
<thead>
<tr>
<th>Variables</th>
<th>Adequate</th>
<th>Poor</th>
</tr>
</thead>
<tbody>
<tr>
<td>Preoperative:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>42.2±12.2</td>
<td>43.1±17.8</td>
</tr>
<tr>
<td>Sex (M:F)</td>
<td>6:4</td>
<td>2:4</td>
</tr>
<tr>
<td>T. bil (mg/dl)*</td>
<td>7.1±8.3</td>
<td>12.7±13.2</td>
</tr>
<tr>
<td>PT (sec)</td>
<td>15.5±2.7</td>
<td>16.0±3.5</td>
</tr>
<tr>
<td>Albumin (g/dl)</td>
<td>2.8±0.6</td>
<td>2.6±0.7</td>
</tr>
<tr>
<td>ICG (20%) (%)</td>
<td>37.7±19.6</td>
<td>28.5±24.0</td>
</tr>
<tr>
<td>LCAT (U)</td>
<td>6.6±5.6</td>
<td>4.1±3.9</td>
</tr>
<tr>
<td>Intraoperative:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Blood loss (U)</td>
<td>12.6±7.9</td>
<td>24.9±28.0</td>
</tr>
</tbody>
</table>

* T. bil, total bilirubin; PT, prothrombin time; ICG (20%), indocyanine green dye retention test at 20 min (normal less than 5%); LCAT, lecithin:cholesterol acyltransferase.
When the elevation of the prothrombin time (over 20 sec in adults and 30 sec in children) is identified immediately after OLTx, we administer fresh frozen plasma to avoid complications related to coagulopathy, such as intracranial or intraabdominal hemorrhage. Introduction of extrinsic coagulation factors, however, may complicate the interpretation of the prothrombin time as an indicator of allograft function.

Plasma LCAT activity, on the other hand, has been well accepted in Japan as an indicator of liver failure following hepatic resection or other surgical procedures for liver disease. LCAT is synthesized in the liver and has a function half-life of 5-6 hr (24), and is not thought to be affected by the administration of fresh frozen plasma. Moreover, it can be measured within about 2 hr without any requirement for unique expertise or equipment. We therefore believe that it may be more useful than current methods—i.e., prothrombin time and coagulation factor levels—for evaluating early hepatic allograft function.

The relationship between the preimplantation histology of the liver allografts and the plasma LCAT activity in the recipient immediately following OLTx suggests that LCAT activity at this time may reflect the condition of the donor liver prior to organ procurement and the degree of preservation or reperfusion injury inflicted upon the allograft.

In summary, plasma LCAT activity assayed immediately following OLTx is a simple, reliable, and inexpensive parameter that may provide a rapid estimation of allograft function and prediction of outcome following OLTx.

TABLE 3. Clinical data of patients with poor graft function after OLTx

<table>
<thead>
<tr>
<th>Case</th>
<th>Age (yr)/Sex</th>
<th>Indication for OLTx</th>
<th>Peak values within 5 days after OLTx</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>70/F</td>
<td>PGNF, status after OLTx for PBC</td>
<td>sGOT (IU/L) 4010; sGPT (IU/L) 4825; PT (sec) 24.2</td>
<td>Died of MOF (3)*</td>
</tr>
<tr>
<td>2.</td>
<td>23/F</td>
<td>PBC</td>
<td>sGOT (IU/L) 2677; sGPT (IU/L) 2902; PT (sec) 23.1</td>
<td>ReTx for rejection (60), doing well</td>
</tr>
<tr>
<td>3.</td>
<td>31/F</td>
<td>PNC (NANB)</td>
<td>sGOT (IU/L) 1818; sGPT (IU/L) 1260; PT (sec) 23.9</td>
<td>Doing well</td>
</tr>
<tr>
<td>4.</td>
<td>61/M</td>
<td>Recurrent HB, status after OLTx</td>
<td>sGOT (IU/L) 6362; sGPT (IU/L) 3603; PT (sec) 31.6</td>
<td>Died of sepsis (18)</td>
</tr>
<tr>
<td>5.</td>
<td>44/F</td>
<td>PNC (alcoholic)</td>
<td>sGOT (IU/L) 2840; sGPT (IU/L) 1660; PT (sec) 24.0</td>
<td>Prolonged respiratory failure, CMV hepatitis, discharged (234), doing well</td>
</tr>
<tr>
<td>6.</td>
<td>55/F</td>
<td>PBC</td>
<td>sGOT (IU/L) 3035; sGPT (IU/L) 2135; PT (sec) 20.2</td>
<td>ReTx for CMV hepatitis (31), died of sepsis (92)</td>
</tr>
<tr>
<td>7.</td>
<td>21/M</td>
<td>Cystic fibrosis</td>
<td>sGOT (IU/L) 400; sGPT (IU/L) 413; PT (sec) 22.7</td>
<td>ReTx for HAT (1), died of pneumonia (15)</td>
</tr>
<tr>
<td>8.</td>
<td>29/F</td>
<td>PGNF, status after OLTx for BC</td>
<td>sGOT (IU/L) 2805; sGPT (IU/L) 1820; PT (sec) 34.2</td>
<td>ReTx for HAT (3), died of MOF (11)</td>
</tr>
<tr>
<td>9.</td>
<td>54/F</td>
<td>PNC (NANB)</td>
<td>sGOT (IU/L) 13,874; sGPT (IU/L) 5106; PT (sec) 46.0</td>
<td>ReTx for PGNF (5), doing well</td>
</tr>
</tbody>
</table>

* Number of postoperative days; OLTx, orthotopic liver transplantation; PT, prothrombin time; PGNF, primary graft nonfunction; MOF, multiple organ failure; PBC, primary biliary cirrhosis; ReTx, retransplantation; PNC, postnecrotic cirrhosis; NANB: non-A-non-B hepatitis; HB, hepatitis B; CMV, cytomegalovirus; HAT, hepatic artery thrombosis; BC, Budd-Chiari syndrome.

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COAGULATION CHANGES FOLLOWING HEPATIC REVASCULARIZATION DURING LIVER TRANSPLANTATION

Paul L. Harper, Roger J. Luddington, Ian Jennings, David Reardon, Muriel J. Seaman, Robin W. Carrell, John R. Klink, Michael Smith, Keith Rolles, and Roy Calne

Departments of Haematology, Anaesthetics, and Surgery, University of Cambridge, Addenbrooke’s Hospital, Cambridge, CB2 2QQ, United Kingdom

The coagulation changes during liver transplantation have been studied in 14 selected patients. Blood usage in all cases was limited to 8.5 liters, and the preoperative coagulation results were only minimally deranged. Bleeding during the operative procedure was easily managed in all cases. Nonetheless, even in this selected group of “low risk” patients, we have demonstrated that during the anhepatic phase and particularly following hepatic revascularization there is activation of both coagulation and fibrinolysis. These findings imply that if bleeding occurs following revascularization, in addition to the use of replacement blood products, treatment should be directed at reducing the consumptive coagulopathy and inhibiting fibrinolysis. We suggest as a first step antithrombin supplementation to maintain activity above 70%, and an antifibrinolytic agent, such as aprotonin, should be considered as adjuncts to therapy at revascularization.