

Pathology of Chronic Rejection: An Overview of Common Findings and Observations About Pathogenic Mechanisms and Possible Prevention

A. J. Demetris, N. Murase, T.E Starzl and J. J. Fung

Definition, Clinical Presentation and Suspected Etiology

Very good to excellent one year patient and graft survival rates at most experienced transplant centers has shifted considerable attention in research away from short term complications like acute rejection to chronic rejection (CR), which is the major obstacle to long-term, morbidity-free survival (reviewed in detail¹). In comparison to acute rejection, chronic rejection is a more indolent, but progressive form of allograft injury, which is largely irreversible and eventually results in allograft failure.

Although the term "chronic" implies a temporally prolonged course, many cases of chronic rejection clearly evolve from severe or inadequately controlled acute rejection episodes because of non-compliance or an inability to maintain immunosuppression, or to monitor the allograft for rejection. Such patients experience a progressive decline in organ function over a period of weeks to months. Another significant cohort of patients slowly develop graft dysfunction and eventual failure over a period of years from chronic rejection. This may be attributable to "clinically silent" rejection episodes that go undetected or to other factors. A third cadre of patients become trapped in a disheartening cycle that alternates between rejection and infection (e.g., CMV, polyoma virus²), in which the immunosuppressive therapy is repeatedly increased then decreased. Each turn of the cycle further damages the organ, which eventually fails with features of chronic rejection.

Three lines of evidence best support the contention that alloimmunity is the major etiologic factor, as originally suspected.^{3,4} First, isografts rarely, if ever, suffer the same set of changes seen in allografts with chronic rejection, although some isografts do develop chronic arterial lesions, but they are delayed in onset and less severe in comparison to allografts.⁵⁻⁸ Secondly, patients who are at high risk of severe or persistent acute rejection;⁹⁻¹⁶ and those inadequately immunosuppressed have a higher rate of chronic rejection.^{17,18} This is likely related to an increased risk of alloreactivity like that seen with donor/recipient racial or ethnic mismatches,¹⁹⁻²³ sex mismatching,^{14,24} viral infections²⁵⁻³² and in patients treated with immune activating drugs.³³ Finally, greater MHC mismatching results in a higher rate of long term

allograft failure, although the data for liver allografts is conflicting.^{19,20,22,23,34-39}

Common Histopathological Features

There are several similar histopathologic and pathophysiologic manifestations of chronic rejection in the various solid organ allografts which enables us to cover the topic from a common perspective. The common features include: a) patchy, organized interstitial inflammation; b) patchy interstitial fibrosis and associated parenchymal atrophy; c) graft vascular disease (GVD) which primarily manifests as fibrointimal hyperplasia of arteries; d) destruction of epithelial-lined conduits; and e) destruction and atrophy of organ-associated lymphoid tissue and lymphatics. Except for widespread GVD, which is rarely detected on biopsy and an uncommon finding in the absence of chronic rejection, none of these features, in isolation, is diagnostic for chronic rejection. Each can be seen with non-chronic rejection related complications. In fact, the usefulness of a biopsy in establishing the diagnosis of chronic rejection is directly related to the ability to sample the target structures.

There are however, certain manifestations of chronic rejection that are accentuated in specific organs because of physioanatomic variations and interactions with the environment. For example, GVD is the primary manifestation of chronic rejection in heart allografts,⁴⁰⁻⁴³ which may be related to the natural tendency of the heart to develop atherosclerosis. In lung transplantation, destruction of the small bronchioles is the major chronic rejection-related insult limiting long-term survival which could be related to bronchial exposure to environmental antigens/infection.^{12,42,44-47} GVD is not a significant problem after lung transplantation.^{12,42,44-46} In the liver, both bile duct loss and GVD (see below) together contribute to allograft failure.⁴⁸⁻⁵² It is clear however that these physioanatomic and "environmental factors" clearly exert their influence in the context of severe and/or persistent alloreactivity, a contention based on the experience with isografts mentioned above.

In addition, there are major differences in the incidence of chronic rejection 5 years after transplantation: it affects roughly 25-50% of hearts; 50-60% isolated lungs, 40-60% pancreas

and 30% of kidney allografts, but only 7–20% of liver allografts (reviewed in greater detail in reference¹). Thus, although there is no accepted generic histopathological definition of chronic rejection equally applicable to all organs, the common features can be used as a general guide for analysis and comparison. Each of these common features is discussed in more detail below.

Patchy, Organized Interstitial Inflammation

A mononuclear interstitial inflammatory infiltrate is a constant feature of chronic rejection, and architecturally organized nodular aggregates of lymphocytes and macrophages often arise from a background of isolated mononuclear cells scattered sparingly throughout the organ.^{1,53,54} Overall, the infiltrates consist primarily of CD4⁺ and CD8⁺ T cells and macrophages.⁵³⁻⁶⁰ In contrast to acute rejection, B cells and plasma cells are seen in greater numbers,^{55,56,61} while eosinophils are less frequently seen.⁶²

In all allografts, the organized nodular aggregates are often located near the adventitia of arteries with GVD, near peripheral nerve trunks, and the serosa or capsule of organs.^{53,54,63} These are also the site of draining lymphatics. The overall arrangement and the occasional presence of germinal centers^{1,53,54} within these nodules is typical of organized lymphoid tissue in the regional lymph nodes, and is indicative of ongoing intra-organ antigen presentation. A similar process occurs in autoimmune diseases, such as Crohn's disease and Hashimoto's thyroiditis.^{53,54}

The aggregates and cells individually scattered throughout the interstitium are often seen in close association with damaged parenchymal cells. Cytokine mRNA analysis and immunohistochemical studies for effector molecules showing granzyme B and the presence of Th1-like cytokines IL-2 and IFN- γ .⁶⁰

Patchy Interstitial Fibrosis and Associated Parenchymal Atrophy

Over time, the necro-inflammatory activity discussed above, results in patchy interstitial fibrosis and atrophy of parenchymal cells. This process is accentuated in adventitia of arteries and in sites of ongoing immunological activity near epithelial-lined conduits (discussed in more detail below). The evolving fibrogenesis is likely related to ongoing parenchymal and microvasculature injury (see below) associated with the release of multiple growth factors,⁶⁴⁻⁶⁸ which results in the deposition of tenascin and other matrix components,^{69,70} endothelin⁷¹ and activation of interstitial myofibroblasts.⁷⁰ The presence of larger scars is indirect evidence that ischemic injury with healed infarcts also contribute to fibrogenesis.⁴⁰

Graft Vascular Disease

Graft vascular disease (GVD) affects the majority of solid organ allografts with chronic rejection to some degree and accounts for the most serious physiological consequences in heart, kidney, and pancreatic allografts. It is of secondary importance in some liver allografts because destruction of the bile ducts is of more importance and GVD is an uncommon problem for lung allografts. Nevertheless, GVD has become synonymous with chronic rejection

and has received the most research attention, even though there are other well-known pathologic aspects of chronic rejection and in some allografts GVD is not a significant problem.

Caveats aside, GVD manifests primarily as concentric arterial intimal thickening and adventitial fibrosis,^{40,41,72-74} but occasional mild venous involvement^{44,74,75} and focal destruction of the microvasculature (e.g. renal glomeruli and peri-biliary capillary plexus)^{48,76,77} are also seen. GVD should be separated from atherosclerosis, which is endemic in the general population, but there are overlapping characteristics that make distinction difficult in some cases. This is particularly true for organs such as the heart that are normally prone to atherosclerosis, but less of a problem in atherosclerotic organs, like the liver.

In general, GVD more often involves *both* the extra-organ (e.g., epicardial, hepatic hilar, etc.) and first and second-order branches of medium-sized intra-organ muscular arteries,^{40,42,72-74} in contrast to atherosclerosis, which primarily affects the extra-organ portion of the vessels. However, involvement of the intra-organ arterial tree by GVD is not as diffuse as one might expect from a review of the pathology literature^{40,42,72-74} and the lesions often begin and evolve more quickly near branch points.⁴⁰

Except for the kidney, arteries commonly affected by GVD are rarely sampled in allograft biopsies routinely used to monitor for rejection. Thus, reconstruction of events preceding the development of GVD depends on examination of many failed allograft and autopsy specimens obtained at serial time points after transplantation. An early phase of GVD with which most investigators are familiar is inflammatory arteritis showing intimal inflammation and/or antibody deposition. This is a histopathological marker of severe acute rejection in all allografts and results in endothelial damage, loss of barrier function and the influx of clotting proteins (including fibrin), platelets, blood cells and lipids,^{65,78-80} all of which disrupt intimal homeostasis and trigger a macrophage influx. The media often shows edema and individual myocyte apoptosis. The adventitia is usually edematous and often contains a cuff of lymphocytes and macrophages. The intimal injury leads to a stereotypic repair response that results in fibrointimal hyperplasia and eventual luminal narrowing.^{64,74,81-86} Intimal inflammation is not observed in milder grades of acute rejection, yet some of these patients go on to develop chronic rejection. Instead, the inflammation is often limited to the adventitia. Even so, adventitial inflammation or injury alone can trigger an arterial repair response culminating in fibrointimal hyperplasia.^{54,87-91} Thus, GVD may not require intimal inflammation or direct endothelial injury from antibodies or cells.

The preferential localization of leukocytes in the adventitia and intima of arteries suggests that these are the most important antigenic targets or sites of arterial damage.^{40,73,83,92-95} That the endothelium is targeted in some reactions is not particularly surprising, since it is well-known to be immunologically active. The adventitia contains a lymphatic plexus which is surrounded by donor dendritic cells making it a site of both peripheral sensitization in acute rejection and a conduit for emigrating leukocytes.^{40,54,83,91,93,95} It is easy to envisage that the shoulder region of a pre-existing donor atherosclerotic lesion would also be highly

immunogenic because of the presence of donor hematolymphoid cells and neovascularization.

As GVD progresses, the intimal and/or adventitial inflammation often persists, but the inflammatory cells migrate to the deeper aspects of the thickened intima where they are separated from an intact endothelial layer by a concentric ring of intimal myofibroblasts. Overall, the developing lesion vaguely resembles the shoulder region of an atherosclerotic plaque, discussed above. The deep intimal focus of inflammatory cells often communicates with an adventitial sheath of lymphocytes and macrophages through a focally disrupted media. Thus, from a morphological perspective, the endothelium no longer appears to be the focus of immunologic activity.

Immunophenotypic analyses have shown that the arterial inflammation consists primarily of an admixture of CD4⁺ and CD8⁺ T cells, some of which show perforin positivity and macrophages, with occasional dendritic cells (marker of ongoing antigenic presentation), eosinophils and plasma cells.^{40,62,83,92,96-98} Mitotic activity within this population occurs at the same time as smooth muscle DNA synthesis, suggesting that the immunological reactions and vessel repair are related.^{1,40} In the final stages of GVD, the media of involved arteries becomes thinned and both the intima and the adventitia thickened, fibrotic and hypocellular.

Focal destruction of the microvasculature of allografts is also part of GVD.^{48,66,76,77} This process is probably related to direct immunological damage to the immunogenic microvascular endothelium and is best observed in renal and liver allografts. The glomerular characteristics evolve through a sequence of changes that eventually result in global glomerulosclerosis.⁷⁷ In the liver, similar changes are seen in the peri-biliary plexus,^{48,76} which shares some morphological and functional similarities with the renal glomerulus.

Destruction of Epithelial-lined Conduits

Epithelial cells that line conduits used for exchange of substances with the environment, such as bronchioles, bile ducts, pancreatic ducts, renal tubular epithelium are particularly prone to damage during chronic rejection. There are several possible non-exclusionary explanations for this observation: 1) the presence of a basement membrane, which could potentially play a role in migration, positioning and co-stimulation of T-cells;⁹⁹ 2) an immunologically active antigenic profile that is significantly different than other parenchymal cells, including class I and II MHC, and various adhesion and co-stimulatory molecules;^{99,100} and 3) the presence of nearby antigen presenting cells and lymphatics that facilitate the functional role of these conduits in processing environmental antigen for local presentation and traffic to the regional lymph nodes.

The bronchioles^{31,100} and bile ducts^{18,48,50,52,101-106} are good examples of the last possible explanation for "targeting" of epithelial conduits. A bronchiocentric or ductulocentric immune response precipitated by environmental or autoantigens in an allograft, creates inflammatory microenvironment that has the potential to trigger a rejection reaction. Either reaction can compromise the structural integrity of the conduit, and focally destroy the

local microvasculature and lymphatic drainage, which in turn can ischemically damage the conduit inhibit efficient antigen clearing. One could easily appreciate how this could lead to a vicious cycle, alternating between a persistent and inadequate response to environmental antigens/infections and allogeneic injury, resulting in a downward spiral of allograft structural integrity and function. In liver and lung allografts, an additional consideration is the exclusive blood supply by the hepatic and bronchial arteries, respectively. Either destruction of these vessels, or failure to revascularize them, can significantly contribute to conduit injury.^{48,49,107}

Destruction and Atrophy of Organ-associated Lymphoid Tissue

All organs contain a network of hematolymphoid cells that travel into and out of, and transiently occupy, the interstitium of all vascularized organs. These cells consist of mature T and B lymphocytes, macrophages, hematopoietic stem cells and dendritic cells, primarily derived from progenitors that migrate hematogenously from the bone marrow, although maturation from local precursors can also contribute to this pool. In concert, they monitor the microenvironment and communicate with regional lymph nodes via the circulation and lymphatics. When the organ becomes an allograft, these cells are called "passenger leukocytes" and they are primarily responsible for triggering acute rejection reactions via the ability of mature dendritic cells to directly stimulate allogeneic T cells.

Organs such as the lungs and intestines have a large specialized compartment of organ or mucosal-associated lymphoid tissue (MALT), commensurate with their task of directly dealing with antigens from the external environment. In contrast, the liver is richly endowed with a large macrophage population, consistent with its role as a filter of various opsonized material and other physiological debris. Although not as extensive or well known, the kidney,¹⁰⁸ heart^{109,110} and pancreas also have considerable intra-organ immune networks.

It is our opinion, that the significance and eventual functional re-establishment of this network has not received enough attention in either acute or chronic rejection, or tolerance induction. In general, the therapeutic window for avoiding graft failure from rejection and infection from over-immunosuppression is most narrow for those organs with the largest component of mature T cells, such as the intestines and lungs. As soon as an allograft is revascularized, recipient immune cells circulate through the organ, including the organ-associated lymphoid tissues (GALT, BALT, portal lymphoid tissue)^{111,115} and regional donor lymph nodes transplanted en bloc, with the organ.¹¹⁶ Donor cells also leave the allograft and lodge in recipient lymphoid tissues.^{117,118} Subsequently a bi-directional "in vivo mixed lymphocyte response" occurs in the recipient lymphoid tissue and in the allograft which manifests as acute rejection.^{111-114,116} The greater component of immunogenic cells, the more robust the initial reaction.

Transplantation of an organ also transiently disrupts the efferent lymphatics resulting in organ edema, which contributes to the re-implantation response. The lymphatic channels reconnect within two to three weeks,^{119,120} unless disrupted by acute rejection, which increases production of lymph fluid and again disrupts the

lymphatic microvasculature. Both of these insults contribute to the reappearance of graft edema and swelling during acute rejection reactions.^{119,121-124}

If immunosuppression is kept high during the early post-transplant period, there is a gradual replacement of donor hematolymphoid cells or immune network with similar recipient cells and the intra-organ network, MALT and regional lymph node architecture is restored.¹¹⁵ However, if the framework is disrupted during acute rejection, repopulation is prevented and the allografts develop chronic rejection.¹¹⁵ In chronic rejection, the lung^{124,125} and intestine¹¹⁴ mucosal-associated lymphoid tissue and regional lymph nodes are often destroyed or undergo atrophy. Patchy interstitial fibrosis also focally disrupts intra-organ lymphatics.^{54,126} Both of these changes undoubtedly contribute to an inability of chronically rejecting allografts to adequately process infectious agents and antigens that are normally cleared via these pathways.^{17,28,31,125,127-129} In fact, it is tempting to speculate that failure to physiologically re-establish these systems after transplantation accounts for the frequent association between infection and chronic rejection. Conversely, one wonders whether this intra-organ immune network also plays a role in maintaining tolerance to the organ.^{54,130-132}

Lastly, disruption of lymphatic drainage, which can occur in the adventitia of arteries can produce arterial injury and intimal thickening similar to that seen with GVD.¹³³ Thus, we⁵⁴ and others before us¹²⁶ have suggested that this potential mechanism of arterial injury might importantly contribute to the development of OA.

The "Special Case" of Liver Allografts

There are two fascinating aspects of chronic liver allograft rejection: its potential reversibility possibly related to ductal regeneration and liver progenitor cells^{150,134} and an appreciably lower incidence than other allografts.^{1,135} Also pertinent is the observation that a liver allograft can also protect other organs from the same donor from chronic rejection.⁵⁴

Theories explaining the special immunological properties of a liver allograft can be broadly separated into two general categories based on whether emphasis is placed on the parenchymal or non-parenchymal fraction. Release of soluble donor MHC class I antigen from the allograft is cited as evidence supporting the importance of the parenchyma.¹³⁶ However, murine liver allografts are routinely accepted between strains of mice that show no difference between the class I loci but are mismatched for class II¹³⁷ and fully allogeneic liver allografts from class I or II MHC deficient mice, which do not shed soluble MHC antigens,^{137,138} are also accepted. Other organs also secrete soluble MHC antigens¹³⁹ but they are routinely rejected. Finally, studies attempting to induce graft acceptance with administration of soluble donor MHC have met with limited success.

Another potential explanation for the importance of the parenchyma relies on the concept that allogeneic hepatocytes provide only one of two signals needed for allogeneic lymphocyte activation,¹⁴⁰ which in turn, could theoretically result in the induction of anergy in the responding lymphocyte populations.¹⁴⁰ Alternatively,

loss of passenger leukocytes from the allograft or graft adaptation could result in ignorance of the allograft.^{142,144} Such a pathway might be especially true for the liver, which is a sink for effete and highly activated cytotoxic T cells.¹⁴³⁻¹⁴⁴

Our focus has been on the donor hematolymphoid cells within the liver, which might initially mediate activation induced clonal purging or deletion¹⁴⁵⁻¹⁵¹ which is followed by long term hematolymphoid microchimerism^{147,152,153} sustained by donor hematopoietic stem contained within the liver.¹⁵⁴⁻¹⁵⁶ An important aspect of both of these mechanisms is that hematolymphoid cells are capable of potent stimulation and have direct access to the recipient lymphoid tissue. However, it is difficult to understand how the initial purging would provide long term protection since one must account for perpetuation of non-reactivity. Freedom from chronic rejection requires long term unresponsiveness. An attractive explanation might combine the initial clonal purging, followed by "ignorance" of the allograft because of the adaptation and replacement of the passenger leukocytes. The situation could be likened to the expression of alloantigens on parenchymal cells in transgenic mice. Unfortunately, this ignorance can be disrupted by viral infections and other local immune activating events,^{142,157,158} which are difficult, if not impossible to avoid in an allograft.

From our perspective, the immunologic mechanisms involved in perpetual graft acceptance and freedom from chronic rejection appear to be active and are probably not different from those required for self-tolerance.^{54,159-162} In essence, to permanently avoid chronic rejection, tolerance to the organ must be induced, and this requires transplantation and functioning of the donor immune system.^{160,161}

Summary

In summary, there are histopathological features of chronic rejection that are common to all solid organ allografts, and immunological injury seems to play a primary role in the initiation and progression of lesions. However, the final phenotypic expression of chronic rejection is dependent on an interaction between immunologic and physiologic/environmental factors that results in one or another of these features predominating in certain organs. Finally, in addition to traditional clinical studies and experimental models, it will be important to study conditions of resistance to chronic rejection in an effort to prevent or avoid this disorder.

References

1. Demetris AJ, Murase N, Lee RG et al. Chronic Rejection. A general overview of histopathology and pathophysiology with emphasis on liver heart and intestinal allografts. *Transplantation Annals* 1997; (in press).
2. Pappo O, Demetris AJ, Raikow RB, Randhawa PS. Human polyoma virus infection of renal allografts: histopathologic diagnosis, clinical significance, and literature review. *Mod Pathol* 1996; 9(2):105-9.
3. Hume DM, Merrill JP, Miller BF, Thorn GW. Experience with renal homotransplantation in the human: Report of nine cases. *J Clin Invest* 1955; 34:327-383.
4. Porter KA, Thomson WB, Owen K, Kenyon JR, Mobray JF, Peart WS. Obliterative vascular changes in four human kidney homotransplants. *British Medical Journal* 1963; 2:639.

5. Veith FJ, Montefusco CM, Blumcke S, Hagstrom JW. Long-term fate of lung autografts charged with providing total pulmonary function. I. Light and electron microscopic studies. *Annals of Surgery* 1979; 190(5):648-53.
6. Norin AJ, Goodell EM, Kamholz SL, Veith FJ, Blumenstock DA. Immunologic, morphologic, and functional evaluation of long-term-surviving beagle lung allograft recipients treated with lethal total-body irradiation, autologous bone marrow, and methotrexate. *Transplantation* 1987; 44(2):179-84.
7. Tullius SG, Heemann U, Hancock WW, Azuma H, Tilney NL. Long-term kidney isografts develop functional and morphologic changes that mimic those of chronic allograft rejection. *Annals of Surgery* 1994; 220(4):425-32.
8. Nadeau KC, Azuma H, Tilney NL. Sequential cytokine dynamics in chronic rejection of rat renal allografts: roles for cytokines RANTES and MCP-1. *PNAS* 1995; 92(19):8729-33.
9. Anonymous. Randomised trial comparing tacrolimus (FK506) and cyclosporin in prevention of liver allograft rejection. European FK506 Multicentre Liver Study Group. *Lancet* 1994; 344(8920):423-8.
10. Hayry P, Mennander A, Yilmaz S et al. Towards understanding the pathophysiology of chronic rejection. *Clinical Investigator* 1992; 70(9):780-90.
11. Matas A. Chronic rejection in renal transplant recipients—risk factors and correlates. *Clinical Transplantation* 1994; 8(3 Pt 2):332-5.
12. Griffith BP, Hardesty RL, Armitage JM et al. A decade of lung transplantation. *Annals of Surgery* 1993; 218(3):310-8.
13. Tejani A, Cortes L, Stablein D. Clinical correlates of chronic rejection in pediatric renal transplantation. A report of the North American Pediatric Renal Transplant Cooperative Study. *Transplantation* 1996; 61(7):1054-8.
14. Candinas D, Gunson BK, Nightingale P, Hubscher S, McMaster P, Neuberger JM. Sex mismatch as a risk factor for chronic rejection of liver allografts. *Lancet* 1995; 346(8983):1117-21.
15. Lee RG, Nakamura K, Tsamandas AC et al. Pathology of human intestinal transplantation [see comments]. *Gastroenterology* 1996; 110(6):1820-34.
16. Wiesner RH, Ludwig J, van Hoek B, Krom RA. Current concepts in cell-mediated hepatic allograft rejection leading to ductopenia and liver failure. *Hepatology* 1991; 14(4 Pt 1):721-9.
17. Whitehead B, Rees P, Sorensen K et al. Incidence of obliterative bronchiolitis after heart-lung transplantation in children. *Journal of Heart & Lung Transplantation* 1993; 12(6 Pt 1):903-8.
18. Panel TW. Terminology for hepatic allograft rejection. *Hepatology* 1995; 22:648-654.
19. Jarcho J, Naftel DC, Shroyer TW et al. Influence of HLA mismatch on rejection after heart transplantation: a multiinstitutional study. The Cardiac Transplant Research Database Group. *Journal of Heart & Lung Transplantation* 1994; 13(4):583-95.
20. Katznelson S, Gjertson DW, Cecka JM. The effect of race and ethnicity on kidney allograft outcome. *Clinical Transplants* 1995:379-94.
21. Devlin JJ, JG OG, Tan KC, Calne RY, Williams R. Ethnic variations in patient and graft survival after liver transplantation. Identification of a new risk factor for chronic allograft rejection. *Transplantation* 1993; 56(6):1381-4.
22. Held PJ, Kahan BD, Hunsicker LG et al. The impact of HLA mismatches on the survival of first cadaveric kidney transplants [see comments]. *New England Journal of Medicine* 1994; 331(12):765-70.
23. Koyama H, Cecka JM, Terasaki PI. Kidney transplants in black recipients. HLA matching and other factors affecting long-term graft survival. *Transplantation* 1994; 57(7):1064-8.
24. Mehra MR, Stapleton DD, Ventura HO et al. Influence of donor and recipient gender on cardiac allograft vasculopathy. An intravascular ultrasound study. *Circulation* 1994; 90(5 Pt 2):II78-82.
25. Grattan MT, Moreno-Cabral CE, Starnes VA, Oyer PE, Stinson EB, Shumway NE. Cytomegalovirus infection is associated with cardiac allograft rejection and atherosclerosis. *JAMA* 1989; 261(24):3561-6.
26. Keenan RJ, Lega ME, Dummer JS et al. Cytomegalovirus serologic status and postoperative infection correlated with risk of developing chronic rejection after pulmonary transplantation. *Transplantation* 1991; 51(2):433-8.
27. Jakel KT, Loning T, Arndt R, Rodiger W. Rejection, herpesvirus infection, and Ki-67 expression in endomyocardial biopsy specimens from heart transplant recipients. *Pathology, Research & Practice* 1992; 188(1-2):27-36.
28. Manez R, White LT, Linden P et al. The influence of HLA matching on cytomegalovirus hepatitis and chronic rejection after liver transplantation. *Transplantation* 1993; 55(5):1067-71.
29. Zeevi A, Pavlick M, Lombardozzi S et al. Immune status of recipients following bone marrow-augmented solid organ transplantation. *Transplantation* 1995; 59(4):616-20.
30. Albar B, Missov E, Serre I, Baldet P, Chaptal PA. Short-term development of transplant-related coronary artery disease in orthotopic cardiac allograft recipients. *Minerva Cardioangiologica* 1995; 43(10):435-8.
31. Siddiqui MT, Garrity ER, Husain AN. Bronchiolitis obliterans organizing pneumonia-like reactions: a nonspecific response or an atypical form of rejection or infection in lung allograft recipients? *Human Pathology* 1996; 27(7):714-9.
32. Lautenschlager I, Hockerstedt K, Jalanko H et al. Persistent Cytomegalovirus In Liver Allografts With Chronic Rejection. *Hepatology* 1997; 25(1):190-194.
33. Feray C, Samuel D, Gigou M et al. An open trial of interferon alfa recombinant for hepatitis C after liver transplantation: antiviral effects and risk of rejection. *Hepatology* 1995; 22(4 Pt 1):1084-9.
34. Doxiadis II, Smits JM, Schreuder GM et al. Association between specific HLA combinations and probability of kidney allograft loss: the taboo concept. *Lancet* 1996; 348(9031):850-3.
35. Kerman RH, Kimball PM, Lindholm A et al. Influence of HLA matching on rejections and short- and long-term primary cadaveric allograft survival. *Transplantation* 1993; 56(5):1242-7.
36. Matas AJ, Burke JF, Jr., DeVault GA, Jr., Monaco A, Pirsch JD. Chronic rejection. [Review] [51 refs]. *Journal of the American Society of Nephrology* 1994; 4(8 Suppl):S23-9.
37. Zhou YC, Cecka JM. Effect of HLA matching on renal transplant survival. *Clinical Transplants* 1993:499-510.
38. Donaldson P, Underhill J, Doherty D et al. Influence of human leukocyte antigen matching on liver allograft survival and rejection: "the dualistic effect". *Hepatology* 1993; 17(6):1008-15.
39. Markus BH, Fung JJ, Gordon RD, Vanek M, Starzl TE, Duquesnoy RJ. HLA histocompatibility and liver transplant survival. *Transplantation Proceedings* 1987; 19(4 Suppl 3):63-5.
40. Demetris AJ, Zerbe T, Banner B. Morphology of solid organ allograft arteriopathy: identification of proliferating intimal cell populations. *Transplantation Proceedings* 1989; 21(4):3667-9.
41. Billingham ME. Pathology and etiology of chronic rejection of the heart. *Clinical Transplantation* 1994; 8(3 Pt 2):289-92.
42. Billingham ME. Pathology of graft vascular disease after heart and heart-lung transplantation and its relationship to obliterative bronchiolitis. [Review] [17 refs]. *Transplantation Proceedings* 1995; 27(3):2013-6.
43. Tazelaar HD, Edwards WD. Pathology of cardiac transplantation: recipient hearts (chronic heart failure) and donor hearts (acute and chronic rejection). *Mayo Clinic Proceedings* 1992; 67(7):685-96.
44. Yousem SA, Paradis IL, Dauber JH et al. Pulmonary arteriosclerosis in long-term human heart-lung transplant recipients. *Transplantation* 1989; 47:564-9.
45. Wahlers T, Haverich A, Schafers HJ et al. Chronic rejection following lung transplantation. Incidence, time pattern and consequences. *European Journal of Cardio Thoracic Surgery* 1993; 7(6):319-23.
46. Sarris GE, Smith JA, Shumway NE et al. Long-term results of combined heart-lung transplantation: the Stanford experience. *Journal of Heart & Lung Transplantation* 1994; 13(6):940-9.
47. Cagle PT, Brown RW, Frost A, Kellar C, Yousem SA. Diagnosis of chronic lung transplant rejection by transbronchial biopsy. *Modern Pathology* 1995; 8(2):137-42.
48. Oguma S, Belle S, Starzl TE, Demetris AJ. A histometric analysis of chronically rejected human liver allografts: insights into the mechanisms of bile duct loss: direct immunologic and ischemic factors. *Hepatology* 1989; 9(2):204-9.
49. Oguma S, Zerbe T, Banner B, Belle S, Starzl TE, Demetris AJ. Chronic liver allograft rejection and obliterative arteriopathy: possible pathogenic mechanisms. *Transplantation Proceedings* 1989; 21(1 Pt 2):2203-7.
50. Freese DK, Snover DC, Sharp HL, Gross CR, Savick SK, Payne WD. Chronic rejection after liver transplantation: a study of clinical, histological and immunological features. *Hepatology* 1991; 13:882-891.

51. Deligeorgi-Politi H, Wight DG, Calne RY, White DG. Chronic rejection of liver transplants revisited [published erratum appears in *Transpl Int* 1995; 8(2):163]. *Transplant International* 1994; 7(6):442-7.
52. Wight DA. Chronic liver transplant rejection: definition and diagnosis. *Transplantation Proceedings* 1996; 28(1):465-7.
53. Luthringer DJ, Yamashita JT, Czer LS, Trento A, Fishbein MC. Nature and significance of epicardial lymphoid infiltrates in cardiac allografts. *Journal of Heart & Lung Transplantation* 1995; 14(3):537-43.
54. Demetris AJ, Murase N, Ye Q et al. An analysis of chronic rejection and obliterative arteriopathy: possible contributions of donor antigen presenting cells and lymphatic disruption. *American Journal of Pathology* 1997; 150:563-578.
55. Demetris AJ, Lasky S, Thiel DHV, Starzl TE, Whiteside T. Induction of DR/IA antigens in human liver allografts: An immunocytochemical and clinicopathologic analysis of twenty failed grafts. *Transplantation* 1985; 40:504-509.
56. Matturri L, Ghidoni P, Palazzi P, Stasi P. Renal allograft rejection: immunohistochemistry of inflammatory cellular subsets and vascular lesions. *Basic & Applied Histochemistry* 1986; 30(2):267-77.
57. McCaughan GW, Davies JS, Waugh JA et al. A quantitative analysis of T lymphocyte populations in human liver allografts undergoing rejection: the use of monoclonal antibodies and double immunolabeling. *Hepatology* 1990; 12(6):1305-13.
58. Stein-Oakley AN, Jablonski P, Tzanidis A et al. Development of chronic injury and nature of interstitial infiltrate in a model of chronic renal allograft rejection. *Transplantation* 1993; 56(6):1299-305.
59. Azuma H, Nadeau KC, Ishibashi M, Tilney NL. Prevention of functional, structural, and molecular changes of chronic rejection of rat renal allografts by a specific macrophage inhibitor. *Transplantation* 1995; 60(12):1577-82.
60. Hayashi M, Martinez OM, Garcia-Kennedy R, So S, Esquivel CO, Krams SM. Expression of cytokines and immune mediators during chronic liver allograft rejection. *Transplantation* 1995; 60(12):1533-8.
61. Winter JB, Clelland C, Gouw AS, Prop J. Distinct phenotypes of infiltrating cells during acute and chronic lung rejection in human heart-lung transplants. *Transplantation* 1995; 59(1):63-9.
62. Nolan CR, Saenz KP, Thomas CA, Murphy KD. Role of the eosinophil in chronic vascular rejection of renal allografts. *American Journal of Kidney Diseases* 1995; 26(4):634-42.
63. Joshi A, Masek MA, B. W. Brown J, Weiss LM, Billingham ME. "Quilty" revisited: a 10-year perspective. *Human Pathology* 1995; 26(5):547-557.
64. Adams DH, Russell ME, Hancock WW, Sayegh MH, Wyner LR, Karnovsky MJ. Chronic rejection in experimental cardiac transplantation: studies in the Lewis-F344 model. *Immunological Reviews* 1993; 134:5-19.
65. Dong C, Redenbach D, Wood S, Battistini B, Wilson JE, McManus BM. The pathogenesis of cardiac allograft vasculopathy. [Review] [54 refs]. *Current Opinion in Cardiology* 1996; 11(2):183-90.
66. Orosz CG. Endothelial activation and chronic allograft rejection. *Clinical Transplantation* 1994; 8(3 Pt 2):299-303.
67. Paul LC, Saito K, Davidoff A, Benediktsson H. Growth factor transcripts in rat renal transplants. *American Journal of Kidney Diseases* 1996; 28(3):441-50.
68. Raines EW, Ross R. Multiple growth factors are associated with lesions of atherosclerosis: specificity or redundancy? [Review] [70 refs]. *Bioessays* 1996; 18(4):271-82.
69. Truong LD, Foster SV, Barrios R et al. Tenascin is an ubiquitous extracellular matrix protein of human renal interstitium in normal and pathologic conditions. *Nephron* 1996; 72(4):579-86.
70. Demirci G, Nashan B, Pichlmayr R. Fibrosis in chronic rejection of human liver allografts: expression patterns of transforming growth factor-TGFbeta1 and TGF-beta3. *Transplantation* 1996; 62(12):1776-83.
71. Forbes RD, Cernacek P, Zheng S, Gomersall M, Guttman RD. Increased endothelin expression in a rat cardiac allograft model of chronic vascular rejection. *Transplantation* 1996; 61(5):791-7.
72. Lin H, Wilson JE, Kendall TJ et al. Comparable proximal and distal severity of intimal thickening and size of epicardial coronary arteries in transplant arteriopathy of human cardiac allografts. *Journal of Heart & Lung Transplantation* 1994; 13(5):824-33.
73. Sibley RK. Morphologic features of chronic rejection in kidney and less commonly transplanted organs. *Clinical Transplantation* 1994; 8(3 Pt 2):293-8.
74. Radio S, Wood S, Wilson J, Lin H, Winters G, McManus B. Allograft vascular disease: comparison of heart and other grafted organs. *Transplantation Proceedings* 1996; 28(1):496-9.
75. Liu G, Butany J, Wanless IR, Cameron R, Greig P, Levy G. The vascular pathology of human hepatic allografts. *Human Pathology* 1993; 24(2):182-8.
76. Matsumoto Y, McCaughan GW, Painter DM, Bishop GA. Evidence that portal tract microvascular destruction precedes bile duct loss in human liver allograft rejection. *Transplantation* 1993; 56(1):69-75.
77. Porter K. Renal Transplantation. In: Heptinstall RH, ed. *Pathology of the Kidney*. 4th ed. Boston: Little, Brown and Company, 1992:1799-1934. vol III).
78. McManus BM, Malcom G, Kendall TJ et al. Lipid overload and proteoglycan expression in chronic rejection of the human transplanted heart. *Clinical Transplantation* 1994; 8(3 Pt 2):336-40.
79. McManus BM, Horley KJ, Wilson JE et al. Prominence of coronary arterial wall lipids in human heart allografts. Implications for pathogenesis of allograft arteriopathy. *American Journal of Pathology* 1995; 147(2):293-308.
80. Lin H, Wilson JE, Roberts CR et al. Biglycan, Decorin, and Versican Protein Expression Patterns In Coronary Arteriopathy Of Human Cardiac Allografts - Distinctness As Compared to Native Atherosclerosis. *Journal of Heart & Lung Transplantation* 1996; 15(12):1233-1247.
81. Cramer DV, Qian S, Harnaha J, Chapman FA, Starzl TE, Makowka L. Accelerated graft arteriosclerosis is enhanced by sensitization of the recipient to donor lymphocytes. *Transplantation Proceedings* 1989; 21(4):3714-5.
82. Cramer DV, Qian SQ, Harnaha J et al. Cardiac transplantation in the rat. I. The effect of histocompatibility differences on graft arteriosclerosis. *Transplantation* 1989; 47(3):414-9.
83. Oguma S, Banner B, Zerbe T, Starzl T, Demetris AJ. Participation of dendritic cells in vascular lesions of chronic rejection of human allografts. *Lancet* 1988; 2(8617):933-6.
84. Paul LC, Hayry P, Foegh M et al. Diagnostic criteria for chronic rejection/accelerated graft atherosclerosis in heart and kidney transplants: joint proposal from the Fourth Alexis Carrel Conference on Chronic Rejection and Accelerated Arteriosclerosis in Transplanted Organs. *Transplantation Proceedings* 1993; 25(2):2022-3.
85. Davies H, al-Tikriti S. Coronary arterial pathology in the transplanted human heart. *International Journal of Cardiology* 1989; 25(1):99-117.
86. Foerster A. Vascular rejection in cardiac transplantation. A morphological study of 25 human cardiac allografts. *Apms* 1992; 100(4):367-76.
87. Chignier E, Eloy R. Adventitial resection of small artery provokes endothelial loss and intimal hyperplasia. *Surg Gynecol Obstet* 1986; 163(4):327-34.
88. Booth RF, Martin JF, Honey AC, Hassall DG, Beesley JE, Moncada S. Rapid development of atherosclerotic lesions in the rabbit carotid artery induced by perivascular manipulation. *Atherosclerosis* 1989; 76(2-3):257-68.
89. Prescott MF, McBride CK, Court M. Development of intimal lesions after leukocyte migration into the vascular wall. *Am J Pathol* 1989; 135(5):835-46.
90. Scott NA, Cipolla GD, Ross CE et al. Identification of a potential role for the adventitia in vascular lesion formation after balloon overstretch injury of porcine coronary arteries. *Circulation* 1996; 93(12):2178-87.
91. Plissonnier D, Nochy D, Poncet P et al. Sequential immunological targeting of chronic experimental arterial allograft. *Transplantation* 1995; 60(5):414-24.
92. Hruban RH, Beschoner WE, Baumgartner WA et al. Accelerated arteriosclerosis in heart transplant recipients is associated with a T-lymphocyte-mediated endothelialitis. *American Journal of Pathology* 1990; 137(4):871-82.
93. Mennander A, Paavonen T, Hayry P. Intimal thickening and medial necrosis in allograft arteriosclerosis (chronic rejection) are independently regulated. *Arteriosclerosis and Thrombosis* 1993; 13(7):1019-25.

94. Goulesbrough DR, Axelsen RA. Arterial endothelialitis in chronic renal allograft rejection: a histopathological and immunocytochemical study [see comments]. *Nephrology, Dialysis, Transplantation* 1994; 9(1):35-40.
95. Russell PS, Chase CM, Winn HJ, Colvin RB. Coronary atherosclerosis in transplanted mouse hearts. I. Time course and immunogenetic and immunopathological considerations. *American Journal of Pathology* 1994; 144(2):260-74.
96. Muller-Hermelink HK, Dammrich JR. [Obliterative transplant vasculopathy: pathogenesis and pathologic mechanisms]. [German]. *Verhandlungen der Deutschen Gesellschaft für Pathologie* 1989; 73:193-206.
97. Vollmer E, Bosse A, Bogenholz J et al. Apolipoproteins and immunohistological differentiation of cells in the arterial wall of kidneys in transplant arteriopathy. Morphological parallels with atherosclerosis. *Pathology, Research & Practice* 1991; 187(8):957-62.
98. Dong C, Wilson JE, Winters GL, McManus BM. Human transplant coronary artery disease: pathological evidence for Fas-mediated apoptotic cytotoxicity in allograft arteriopathy. *Laboratory Investigation* 1996; 74(5):921-31.
99. Demetris AJ, ed. *Immunopathology of the Human Biliary Tree*. Boca Raton: CRC Press, 1996. (Sirica AE, ed. *Pathobiology of the Biliary Tree*; vol (in press)).
100. Mauck KA, Hosenpud JD. The bronchial epithelium: a potential allogeneic target for chronic rejection after lung transplantation. *Journal of Heart & Lung Transplantation* 1996; 15(7):709-14.
101. Fennell RH. Ductular damage in liver transplant rejection: its similarity to that of primary biliary cirrhosis and graft-versus-host disease. *Pathol Annu* 1981; 1981:289-294.
102. Portmann B, Neuberger J, Williams R. Intrahepatic bile duct lesions. In: Calne RY, ed. *Liver Transplantation. the Cambridge-Kings College Hospital Experience*. London: Grune & Stratton, 1983:279.
103. Grond J, Gouw AS, Poppema S, Sloof MJH, Gips CH. Chronic rejection in liver transplants: a histopathologic analysis of failed grafts and antecedent serial biopsies. *Transplantation Proceedings* 1986; 18:128-135.
104. Vierling JM, R. H. Fennell J. Histopathology of early and late human hepatic allograft rejection. Evidence of progressive destruction of interlobular bile ducts. *Hepatology* 1985; 4:1076-1082.
105. Hoek BV, Wiesner R, Krom R, Ludwig J, Moore S. Severe ductopenic rejection following liver transplantation: incidence, time of onset, risk factors, treatment and outcome. *Seminars in Liver Disease* 1992; 12:41-50.
106. Lowes J, Hubscher S, Neuberger J. Chronic rejection of the liver allograft. *Gastroenterology Clinics of North America* 1993; 22:401-420.
107. Baudet EM, Dromer C, Dubrez J et al. Intermediate-term results after en bloc double-lung transplantation with bronchial arterial revascularization. *Bordeaux Lung and Heart-Lung Transplant Group. Journal of Thoracic & Cardiovascular Surgery* 1996; 112(5):1292-9.
108. Kaissling B, Hegyi I, Löffing J, Le Hir M. Morphology of interstitial cells in the healthy kidney. *Anat Embryol (Berl)* 1996; 193(4):303-18.
109. Austyn JM, Hankins DF, Larsen CP, Morris PJ, Rao AS, Roake JA. Isolation and characterization of dendritic cells from mouse heart and kidney. *J Immunol* 1994; 152(5):2401-10.
110. Holzinger C, Zuckermann A, Reinwald C et al. Are T cells from healthy heart really only passengers? Characterization of cardiac tissue T cells. *Immunol Lett* 1996; 53(2-3):63-7.
111. Prop J, Kuipers K, Petersen AH, Bartels HL, Nieuwenhuis P, Wildevuur CR. Why are lung allografts more vigorously rejected than hearts? *Journal of Heart Transplantation* 1985; 4(4):433-6.
112. Prop J, Wildevuur CR, Nieuwenhuis P. Lung allograft rejection in the rat. II. Specific immunological properties of lung grafts. *Transplantation* 1985; 40(2):126-31.
113. Prop J, Nieuwenhuis P, Wildevuur CR. Lung allograft rejection in the rat. I. Accelerated rejection caused by graft lymphocytes. *Transplantation* 1985; 40(1):25-30.
114. Murase N, Demetris AJ, Kim DG, Todo S, Fung JJ, Starzl TE. Rejection of multivisceral allografts in rats: a sequential analysis with comparison to isolated orthotopic small-bowel and liver grafts. *Surgery* 1990; 108(5):880-9.
115. Murase N, Demetris AJ, Matsuzaki T et al. Long survival in rats after multivisceral versus isolated small-bowel allotransplantation under FK 506. *Surgery* 1991; 110(1):87-98.
116. Fung J, Zeevi A, Demetris AJ et al. Origin of lymph node derived lymphocytes in human hepatic allografts. *Clin Transplant* 1989; 3:316-324.
117. Larsen CP, Morris PJ, Austyn JM. Migration of dendritic leukocytes from cardiac allografts into host spleens. A novel route for initiation of rejection. *J Exp Med* 1990; 171:307-314.
118. Demetris A, Qian S, H Sun ea. Early events in liver allograft rejection: delineation of sites of simultaneous intragraft and recipient lymphoid tissue sensitization. *American Journal of Pathology* 1991; 138:609.
119. Malek P, Vrabel J, Kolc J. Lymphatic aspects of experimental and clinical renal transplantation. *Bulletin de la Societe Internationale de Chirurgie* 1969; 28(1):110-4.
120. Kocandrie V, Houttuin E, Prohaska JV. Regeneration of the lymphatics after autotransplantation and homotransplantation of the entire small intestine. *Surgery, Gynecology & Obstetrics* 1966; 122(3):587-92.
121. Malek P, Vrabel J. Lymphatic system and organ transplantation. *Lymphology* 1968; 1(1):4-22.
122. Cockett AT, Sakai A, Netto IC. Kidney lymphatics: an important network in transplantation. *Transactions of the American Association of Genito Urinary Surgeons* 1973; 65:73-6.
123. Eliska O, Eliskova M, Mirejovsky P. Lymph vessels of the transplanted kidney. *Nephron* 1986; 44(2):136-41.
124. Ruggiero R, Fietsam R, Jr., Thomas GA et al. Detection of canine allograft lung rejection by pulmonary lymphoscintigraphy. *Journal of Thoracic & Cardiovascular Surgery* 1994; 108(2):253-8.
125. Hruban RH, Beschoner WE, Baumgartner WA et al. Depletion of bronchus-associated lymphoid tissue associated with lung allograft rejection. *American Journal of Pathology* 1988; 132(1):6-11.
126. Kline IK, Thomas PA. Canine lung allograft lymphatic alterations. *Annals of Thoracic Surgery* 1976; 21(6):532-5.
127. Durham JR, Nakhleh RE, Levine A, Levine TB. Persistence of interstitial inflammation after episodes of cardiac rejection associated with systemic infection. *Journal of Heart & Lung Transplantation* 1995; 14(4):774-80.
128. Heemann UW, Tullius SG, Schmid C, Philipp T, Tilney NL. Infection-associated cellular activation accelerates chronic renal allograft rejection in rats. *Transplant International* 1996; 9(2):137-40.
129. Wallwork J. Risk factors for chronic rejection in heart and lungs—why do hearts and lungs rot? *Clinical Transplantation* 1994; 8(3 Pt 2):341-4.
130. Saleem M, Sawyer GJ, Schofield RA, Seymour ND, Gustafsson K, Fabre JW. Discordant expression of major histocompatibility complex class II antigens and invariant chain in interstitial dendritic cells. Implications for self-tolerance and immunity. *Transplantation* 1997; 63(8):1134-8.
131. Steinman RM, Pack M, Inaba K. Dendritic cells in the T-cell areas of lymphoid organs. *Immunol Rev* 1997; 156:25-37.
132. Inaba K, Pack M, Inaba M, Sakuta H, Isdell F, Steinman RM. High levels of a major histocompatibility complex II-self peptide complex on dendritic cells from the T cell areas of lymph nodes. *J Exp Med* 1997; 186(5):665-72.
133. Solti F, Jellinek H, Schneider F, Lengyel E, Berczi V, Kekesi V. Lymphatic arteriopathy: damage to the wall of the canine femoral artery after lymphatic blockade. *Lymphology* 1991; 24(2):54-9.
134. Hubscher SG, Neuberger JM, Buckels JAC, Elias E, McMaster P. Vanishing bile-duct syndrome after liver transplantation - is it reversible? *Transplantation* 1991; 51:1004-1110.
135. Demetris AJ, Murase N, Delaney CP, Woan M, Fung JJ, Starzl TE. The liver allograft, chronic (ductopenic) rejection, and microchimerism: What can they teach us? *Transplantation Proceedings* 1995; 27:67-70.
136. Davies HS, Pollard SG, Calne RY. Soluble HLA antigens in the circulation of liver graft recipients. *Transplantation* 1989; 47:524-527.
137. Qian S, Sun H, Demetris AJ, Fu F, Starzl TE, Fung JJ. Liver graft induced donor specific unresponsiveness without class I and/or class II antigen differences. *Transplantation Proceedings* 1993; 25(1 Pt 1):362-3.
138. Qian S, Demetris AJ, Murase N, Rao AS, Fung JJ, Starzl TE. Murine liver allograft transplantation: tolerance and donor cell chimerism. *Hepatology* 1994; 19:916-924.

- I39. Rhynes VK, McDonald JC, Gelder FB et al. Soluble HLA class I in the serum of transplant recipients. *Annals of Surgery* 1993; 217(5):485-9.
- I40. Matzinger P. Tolerance, danger, and the extended family. *Annual Review of Immunology* 1994; 12:991-1045.
- I41. Slattery RM, Miller JF, Heath WR, Charlton B. Failure of a protective major histocompatibility complex class II molecule to delete autoreactive T cells in autoimmune diabetes. *Proceedings of the National Academy of Sciences of the United States of America* 1993; 90(22):10808-10.
- I42. Miller JF, Heath WR. Self-ignorance in the peripheral T-cell pool. *Immunological Reviews* 1993; 133:131-50.
- I43. Bertolino P, Heath WR, Hardy CL, Morahan G, Miller JF. Peripheral deletion of autoreactive CD8+ T cells in transgenic mice expressing H-2Kb in the liver. *European Journal of Immunology* 1995; 25(7):1932-42.
- I44. Kamada N. The immunology of experimental liver transplantation in the rat. *Immunology* 1985; 55:369-389.
- I45. Starzl TE. *Host-Graft Adaptation. Experience in Renal Transplantation.* Philadelphia PA: W. B. Saunders Company, 1964:164-170.
- I46. Starzl TE, Marchioro TL, Waddell WR. The reversal of rejection in human renal homografts with subsequent development of homograft tolerance. *Surg Gynecol Obstet* 1963; 117:385-395.
- I47. Starzl TE, Demetris AJ, Murase N, Ildstad S, Ricordi C, Trucco M. Cell migration, chimerism, and graft acceptance [see comments]. *Lancet* 1992; 339(8809):1579-82.
- I48. Sun J, McCaughan GW, Matsumoto Y, Sheil AG, Gallagher ND, Bishop GA. Tolerance to rat liver allografts. I. Differences between tolerance and rejection are more marked in the B cell compared with the T cell or cytokine response. *Transplantation* 1994; 57(9):1349-57.
- I49. Sun J, Sheil R, McCaughan G, Jung S, Gallagher N, Bishop G. Strong tolerance mediated by allografting in the rat is due to donor liver leukocytes. *Transplantation Proceedings* 1995; 27(6):3578.
- I50. Sun J, McCaughan GW, Gallagher ND, Sheil AG, Bishop GA. Deletion of spontaneous rat liver allograft acceptance by donor irradiation. *Transplantation* 1995; 60(3):233-6.
- I51. Bishop GA, Sun J, DeCruz DJ et al. Tolerance to rat liver allografts. III. Donor cell migration and tolerance-associated cytokine production in peripheral lymphoid tissues. *Journal of Immunology* 1996; 156(12):4925-31.
- I52. Starzl TE, Demetris AJ, Trucco M et al. Cell migration and chimerism after whole-organ transplantation: the basis of graft acceptance [see comments]. *Hepatology* 1993; 17(6):1127-52.
- I53. Starzl TE, Demetris AJ. Transplantation Milestones: viewed with one- and two-way paradigms of tolerance. *JAMA* 1995; 273:876-879.
- I54. Murase N, Starzl TE, Ye Q et al. Multilineage hematopoietic reconstitution of supralethally irradiated rats by syngeneic whole organ transplantation: with particular reference to the liver. *Transplantation* 1996; 61:1-4.
- I55. Taniguchi H, Toyoshima T, Fukao K, Nakauchi H. Presence of hematopoietic stem cells in the adult liver. *Nature Medicine* 1996; 2:198-203.
- I56. Hays EF, Hays DM, Golde DW. Hematopoietic stem cells in mouse liver. *Exp Hematol* 1978; 6:18.
- I57. Heath WR, Karamalis F, Donoghue J, Miller JF. Autoimmunity caused by ignorant CD8+ T cells is transient and depends on avidity. *Journal of Immunology* 1995; 155(5):2339-49.
- I58. Nossal GJ, Herold KC, Goodnow CC. Autoimmune tolerance and type I (insulin-dependent) diabetes mellitus. *Diabetologia* 1992; 35 Suppl 2:S49-59.
- I59. Liegeois A, Escourrou J, Ouvre E, Charriere J. Microchimerism: a stable state of low-ratio proliferation of allogeneic bone marrow. *Transplantation Proceedings* 1977; 9:273-6.
- I60. Coutinho A. Beyond clonal selection and network. *Immuno Rev* 1989; 110:63-87.
- I61. Demetris AJ, Murase N, Rao AS, Starzl TE. The role of passenger leukocytes in rejection and "tolerance" after solid organ transplantation: a potential explanation of a paradox. *Rejection and Tolerance.* J. L. Touraine et al. ed. Netherlands: Kluwer Academic Publishers, 1994:325-392.
- I62. Delaney CP, Murase N, Chen-Woan M et al. Allogeneic hematolymphoid microchimerism and prevention of autoimmune disease in the rat: a relationship between allo- and autoimmunity. *The Journal of Clinical Investigation* 1996; 97:217-225.

Supported by NIH 1 RO1DK49615-01, NIH 1 RO1 AI 38899-01A2 and 1 RO1 AI40329-02(DK)